On the mechanism of biosynthesis of leukotrienes and related compounds

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[10D-³H; 3-¹⁴C]- and [10L-³H; 3-¹⁴C]arachidonic acids were incubated with human polymorphonuclear leukocytes and with human platelets. Leukotriene B₄ and 5(S),12(S)-dihydroxy-6trans,8cis,10trans,14-cis-eicosatetraenoic acid (5,12-DHETE) were isolated and the ³H/¹⁴C ratios determined. It could be concluded that the 10D (pro-R)-hydrogen is eliminated in the conversion of 5(S)-hydroperoxy-6trans,8-cis,11cis,14cis-eicosatetraenoic acid into leukotriene A₄ whereas in the conversion of arachidonic acid into 5,12-DHETE the 10L (pro-S)-hydrogen is lost. Incubation of the doubly labeled arachidonic acids with human platelets confirmed and extended previous data on the stereochemistry of the hydrogen removal from C-10 during the conversion into 12(S)-hydroperoxy-5cis,8cis,10trans,14cis-eicosatetraenoic acid, i.e., the 10L (pro-S)-hydrogen is eliminated and the 10D (pro-R)-hydrogen retained.

Leukotriene A₄ 5(S),12(S)-dihydroxy-6,8,10,14-eicosatetraenoic acid
12(S)-hydroperoxy-5,8,10,14-eicosatetraenoic acid
Stereospecific hydrogen removal Isotope effect

1. INTRODUCTION

Two reactions are involved in the formation of leukotriene A_4 (LTA₄) from arachidonic acid:

- (1) A lipoxygenase reaction by which arachidonic acid is transformed into 5(S)-hydroperoxy-6trans,8cis,11cis,14cis-eicosatetraenoic acid (5-HPETE) [1];
- (2) A dehydrase reaction in which the hydroperoxide is cyclized into 5(S)-trans-5,6-oxido-7trans,9trans,11cis,14cis-eicosatetraenoic acid (LTA₄) [2].

Several compounds are formed by further transformation of LTA₄; i.e., leukotriene B₄ (LTB₄, 5(S),12(R)-dihydroxy-6cis,8trans,10trans,14cis-eicosatetraenoic acid), 5(S),12(R)-dihydroxy-6trans, 8trans, 10trans, 14cis-eicosatetraenoic acid, and 5(S),12(S)-dihydroxy-6trans,8trans,10trans,14cis-eicosatetraenoic acid as well as the amino acid containing leukotrienes LTC₄, LTD₄, and LTE₄ [2]. Human polymorphonuclear leukocytes have also

been found to produce 5(S),12(S)-dihydroxy-6-trans,8cis,10trans,14cis-eicosatetraenoic acid (5,12-DHETE) [3,4]. This dihydroxy acid, although isomeric with LTB₄, is not formed from LTA₄ but by double dioxygenation of arachidonic acid. The compound is therefore not included in the leukotriene family.

This work is concerned with the stereochemistry of the hydrogen removal from C-10 of arachidonic acid during the biosynthesis of leukotrienes and of 5,12-DHETE.

2. MATERIALS AND METHODS

Human polymorphonuclear leukocytes (HPMNL) were isolated from leukocyte concentrates obtained from blood as in [5]. [$10L^{-3}H$; $3^{-14}C$]arachidonic acid was prepared as in [6] (see fig. 1). [$10D^{-3}H$; $3^{-14}C$]arachidonic acid was obtained in a similar way except for the use of (+) α -phenylethylamine for preparation of 3L-hydroxytridecanoic acid (cf. [7]; see fig. 1). The yield of labeled arachidonic acids from labeled

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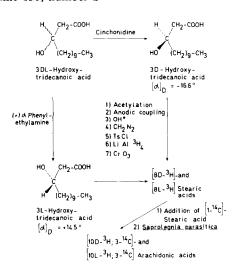


Fig. 1. Reactions used to prepare [10D-³H; 3-¹⁴C]- and [10L-³H; 3-¹⁴C]arachidonic acids; 'Ts', p-toluenesulfonyl.

stearic acids following incubation with the fungus, Saprolegnia parasitica, was 0.5-2%.

The cell preparation ($\sim 100 \times 10^6 \text{ HPMNL/ml}$; contaminated with platelets) was stirred for 10 min at 37°C in the presence of ionophore A 23187 (5 µM) and the doubly labeled arachidonic acids $(150 \,\mu\text{M}, 0.05-0.5 \,\mu\text{Ci of}^{-14}\text{C})$. The incubations were stopped by the addition of 1.5 vol. of methanol and the diethyl ether extracts were subjected to silicic acid chromatography (column, 1 g of silicic acid CC-4 obtained from Mallinckrodt). Elution was performed stepwise with diethyl ether-hexane 1:9 (v/v), diethyl ether-hexane 4:6 (v/v), and ethyl acetate. The material eluted with ethyl acetate was subjected to reversed-phase and straight-phase high-performance liquid chromatography essentially as in [3]. Here, the methyl esters of 5.12-DHETE and LTB4 were identified and collected.

The doubly-labeled arachidonic acids were also incubated with suspensions of human platelets [6]. The products, i.e., 12-HETE, 12-HHT and thromboxane B₂ (TXB₂) [8] were isolated in form of their methyl esters by thin-layer chromatography.

³H/¹⁴C ratios of the incubated arachidonic acids as well as of the products formed in leukocytes and platelets were determined with a Packard TriCarb model 3375 liquid scintillation spectrometer using Instagel® as scintillation fluor.

Table 1

Relative retention of ³H in LTB₄ and 5,12-DHETE observed upon incubation of [10D-³H; 3-¹⁴C]- and

[10L-3H: 3-14Clarachidonic acids with HPMNL

20:4 incubated ³ H/ ¹⁴ C (%)	LTB_4 $^3H/^{14}C$ (%)	5,12-DHETE ³ H/ ¹⁴ C (%)	
[10D- ³ H; 3- ¹⁴ C]20:4			
100	22	128	
100	27	149	
100	_	139	
[10L- ³ H; 3- ¹⁴ C]20:4			
100	98	9	

3. RESULTS AND DISCUSSION

3.1. Incubation with leukocytes

[10D-³H; 3-¹⁴C]- and [10L-³H; 3-¹⁴C]arachidonic acids were incubated with suspensions of HPMNL as above. ³H/¹⁴C ratios of 5,12-DHETE and LTB₄ relative to that of the corresponding precursor acid are given in table 1.

As seen, during the conversion of the 10D-tritio arachidonic acid into LTB₄ tritium was largely lost. On the other hand, there was no loss of ³H during the formation of 5,12-DHETE. Instead a certain enrichment of tritium was observed (table 1). LTB₄ formed from the 10L-tritio arachidonic acid retained the ³H label whereas 5,12-DHETE lost most of the tritium.

These data show that the 10D (pro-R)-hydrogen is lost during the formation of LTA4 from 5-HPETE whereas the 10L (pro-S)-hydrogen is lost upon formation of 5,12-DHETE from arachidonic acid. The enrichment of tritium in 5,12-DHETE observed when formed from [10D-3H; 3-14C]arachidonic acid suggests the presence of isotope effects in the conversions of the 10D-tritio arachidonic acid. A likely explanation for the enrichment involves the presence of an isotope effect in the conversion of [10D-3H; 3-14C]5-HPETE into [3-14C]LTA₄. 5-HPETE remaining unconverted will thus be enriched with respect to tritium. Dioxygenation at C-12 does not involve elimination of the 10D-tritium ([6]; table 2). Therefore the resulting [10-3H; 3-14C]5,12-DHETE should be enriched with tritium. In order to study this question

Table 2

Relative retention of ³H in 12-HETE, 12-HHT and TXB₂ observed following incubation of [10D-³H; 3-¹⁴C]-and [10L-³H; 3-¹⁴C]arachidonic acids with human platelets

20:4 incubated ³ H/ ¹⁴ C (%)	12-HETE ³ H/ ¹⁴ C (%)	12-HHT ³ H/ ¹⁴ C (%)	TXB ₂ ³ H/ ¹⁴ C (%)
[10D- ³ H; 3- ¹⁴ C]20:4 100 100 ^a	95 96	2	98
[10L- ³ H; 3- ¹⁴ C]20:4 100 ^a	9		-

^aIndomethacin (10 µg/ml) was added

Fig. 2. Scheme of transformation of arachidonic acid into LTA₄ and 5,12-DHETE; (*) 10D (*pro-R*)-hydrogen of arachidonic acid and 5-HPETE.

in more detail, a separate experiment was carried out in which 5,12-DHETE, as well as 5-HETE were isolated and analysed following incubation of [10D-3H; 3-14C]arachidonic acid. In agreement with the interpretation discussed above (fig. 2) 5,12-DHETE as well as 5-HETE (reduction product of 5-HPETE) were found to be enriched with respect to tritium (130% and 167% relative to pre-

cursor, respectively). It thus appears that 5,12-DHETE may be formed by the sequence arachidonic acid \rightarrow 5-HPETE \rightarrow 5,12-DHETE (fig. 2). However, these data do not exclude the possibility of simultaneous formation of 5,12-DHETE by the alternate sequence of reactions; i.e., arachidonic acid \rightarrow 12-HPETE \rightarrow 5,12-DHETE.

3.2. Incubation with platelets

[10D-³H; 3-¹⁴C]- and [10L-³H; 3-¹⁴C]arachidonic acids were incubated with suspensions of human platelets as in [6,8]. Table 2 gives the relative retentions of ³H in the products.

It has been found that tritium is lost during the conversion of [10L-3H; 3-14C]arachidonic acid into 12-HETE [6]. This result was confirmed here. Furthermore, the 10D-tritio arachidonic acid was found to retain its ³H label upon conversion into 12-HETE (table 2). Also, as would be expected [8], TXB₂ retained the ³H label when formed from [10D-3H; 3-14C]arachidonic acid, whereas 12-HHT was essentially devoid of ³H (table 2).

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